

YESCARTA® (axicabtagene ciloleucel) suspension for intravenous infusion

PACKAGE INSERT

WARNING: CYTOKINE RELEASE SYNDROME and NEUROLOGIC TOXICITIES

- **Cytokine Release Syndrome (CRS), including fatal or life-threatening reactions, occurred in patients receiving YESCARTA. Do not administer YESCARTA to patients with active infection or inflammatory disorders, or active graft-versus-host disease (GVHD). Treat severe or life-threatening CRS with tocilizumab or tocilizumab and corticosteroids [see Dosage and Administration (2.2, 2.3), Warnings and Precautions (5.1)].**
- **Neurologic toxicities, including fatal or life-threatening reactions, occurred in patients receiving YESCARTA, including concurrently with CRS or after CRS resolution. Monitor for neurologic toxicities after treatment with YESCARTA. Provide supportive care and/or corticosteroids as needed [see Dosage and Administration (2.2, 2.3), Warnings and Precautions (5.2)].**
- **YESCARTA is available only through a Controlled Distribution Program [see Warnings and Precautions (5.3)].**

1 INDICATIONS AND USAGE

YESCARTA is a CD19-directed genetically modified autologous T cell immunotherapy indicated for the treatment of:

1.1 Large B-cell Lymphoma

- Adult patients with diffuse large B-cell lymphoma (DLBCL) and high-grade B-cell lymphoma (HGBL) that relapses within 12 months from completion of, or is refractory to, first-line chemoimmunotherapy.
- Adult patients with relapsed or refractory large B-cell lymphoma (LBCL) after two or more lines of systemic therapy, including DLBCL not otherwise specified, primary mediastinal large B-cell lymphoma (PMBCL), HGBL, and DLBCL arising from follicular lymphoma.

Limitations of Use: YESCARTA is not indicated for the treatment of patients with primary central nervous system lymphoma.

2 DOSAGE AND ADMINISTRATION

For autologous use only. For intravenous use only.

2.1 Dose

Each single infusion bag of YESCARTA contains a suspension of chimeric antigen receptor (CAR)-positive T cells in approximately 68 mL. The target dose is 2×10^6 CAR-positive viable T cells per kg body weight, with a maximum of 2×10^8 CAR-positive viable T cells.

2.2 Administration

YESCARTA is for autologous use only. The patient's identity must match the patient identifiers on the YESCARTA cassette and infusion bag. Do not infuse YESCARTA if the information on the patient-specific label does not match the intended patient.

Preparing Patient for YESCARTA Infusion

Confirm availability of YESCARTA prior to starting the lymphodepleting regimen.

Pre-treatment

- Administer a lymphodepleting chemotherapy regimen of cyclophosphamide 500 mg/m² intravenously and fludarabine 30 mg/m² intravenously on the fifth, fourth, and third day before infusion of YESCARTA.

Premedication

- Administer acetaminophen/paracetamol and diphenhydramine approximately 1 hour before YESCARTA infusion.
- Consider the use of prophylactic corticosteroid in patients after weighing the potential benefits and risks [*see Warnings and Precautions (5.1 and 5.2)*].

Preparation of YESCARTA for Infusion

Coordinate the timing of YESCARTA thaw and infusion. Confirm the infusion time in advance and adjust the start time of YESCARTA thaw such that it will be available for infusion when the patient is ready.

- Confirm patient identity: Prior to YESCARTA preparation, match the patient's identity with the patient identifiers on the YESCARTA cassette.
- Do not remove the YESCARTA product bag from the cassette if the information on the patient-specific label does not match the intended patient.
- Once patient identification is confirmed, remove the YESCARTA product bag from the cassette and check that the patient information on the cassette label matches the bag label.
- Inspect the product bag for any breaches of container integrity such as breaks or cracks before thawing. If the bag is compromised, follow the local guidelines (or contact Kite).
- Place the infusion bag inside a second sterile bag per local guidelines.
- Thaw YESCARTA at approximately 37°C using either a water bath or dry thaw method until there is no visible ice in the infusion bag. Gently mix the contents of the bag to disperse clumps of cellular material. If visible cell clumps remain continue to gently mix the contents of the bag. Small clumps of cellular material should disperse with gentle manual mixing. Do not wash, spin down, and/or re-suspend YESCARTA in new medium prior to infusion.
- Once thawed, YESCARTA may be stored at room temperature (20°C to 25°C) for up to 3 hours.

Administration

- For autologous use only.
- Ensure that tocilizumab and emergency equipment are available prior to infusion and during the recovery period.
- Do NOT use a leukodepleting filter.
- Central venous access is recommended for the infusion of YESCARTA.
- Confirm the patient's identity matches the patient identifiers on the YESCARTA product bag.
- Prime the tubing with normal saline prior to infusion.
- Infuse the entire contents of the YESCARTA bag within 30 minutes by either gravity or a peristaltic pump. YESCARTA is stable at room temperature for up to 3 hours after thaw.
- Gently agitate the product bag during YESCARTA infusion to prevent cell clumping.
- After the entire content of the product bag is infused, rinse the tubing with normal saline at the same infusion rate to ensure all product is delivered.

YESCARTA contains human blood cells that are genetically modified with replication incompetent retroviral vector. Follow universal precautions and local biosafety guidelines for handling and disposal to avoid potential transmission of infectious diseases.

Monitoring

- Administer YESCARTA at a certified healthcare facility.
- Monitor patients at least daily for 7 days at the certified healthcare facility following infusion for signs and symptoms of cytokine release syndrome (CRS) and neurologic toxicities.

- Instruct patients to remain within proximity of the certified healthcare facility for at least 4 weeks following infusion.

2.3 Management of Severe Adverse Reactions

Cytokine Release Syndrome (CRS)

Identify CRS based on clinical presentation [*see Warnings and Precautions (5.1)*]. Evaluate for and treat other causes of fever, hypoxia, and hypotension. If CRS is suspected, manage according to the recommendations in Table 1. Patients who experience Grade 2 or higher CRS (e.g., hypotension not responsive to fluids, or hypoxia requiring supplemental oxygenation) should be monitored with continuous cardiac telemetry and pulse oximetry. For patients experiencing severe CRS, consider performing an echocardiogram to assess cardiac function. For severe or life-threatening CRS, consider intensive-care supportive therapy.

Table 1. CRS Grading and Management Guidance

CRS Grade^a	Tocilizumab	Corticosteroids
Grade 1 Symptoms require symptomatic treatment only (e.g., fever, nausea, fatigue, headache, myalgia, malaise).	If symptoms (e.g., fever) not improving after 24 hours, consider managing as Grade 2.	If not improving after 3 days, administer one dose of dexamethasone 10 mg intravenously.
Grade 2 Symptoms require and respond to moderate intervention. Oxygen requirement less than 40% FiO ₂ or hypotension responsive to fluids or low-dose of one vasopressor or Grade 2 organ toxicity. ^b	Administer tocilizumab ^c 8 mg/kg intravenously over 1 hour (not to exceed 800 mg). If no clinical improvement in the signs and symptoms of CRS after the first dose, repeat tocilizumab every 8 hours as needed. Limit to a maximum of 3 doses in a 24-hour period; maximum total of 4 doses. If improving, discontinue tocilizumab.	Administer dexamethasone 10 mg intravenously once daily. If improving, manage as Grade 1 above and continue corticosteroids until the severity is Grade 1 or less, then quickly taper as clinically appropriate. If not improving, manage as appropriate grade below.
Grade 3 Symptoms require and respond to aggressive intervention. Oxygen requirement greater than or equal to 40% FiO ₂ or hypotension requiring high-dose or multiple vasopressors or Grade 3 organ toxicity or Grade 4 transaminitis.	Per Grade 2. If improving, manage as appropriate grade above.	Dexamethasone 10 mg intravenously 3 times a day. If improving, manage as appropriate grade above and continue corticosteroids until the severity is Grade 1 or less, then quickly taper as clinically appropriate. If not improving, manage as Grade 4.
Grade 4 Life-threatening symptoms. Requirements for ventilator support, continuous veno-venous hemodialysis (CVVHD) or Grade 4 organ toxicity (excluding transaminitis).	Per Grade 2. If improving, manage as appropriate grade above.	Administer methylprednisolone 1000 mg intravenously once per day for 3 days. If improving, manage as appropriate grade above and continue corticosteroids until the severity is Grade 1 or less, then taper as clinically appropriate. If not improving, consider methylprednisolone 1000 mg 2-3 times a day or alternate therapy. ^d

a. Lee et al. 2014.

b. Refer to Table 2 for management of neurologic toxicity.

c. Refer to tocilizumab package insert for details.

d. Alternate therapy includes (but is not limited to): anakinra, siltuximab, ruxolitinib, cyclophosphamide, Intravenous Immune Globulin (IVIG) and Anti-Thymocyte Globulin (ATG).

Neurologic Toxicity

Monitor patients for signs and symptoms of neurologic toxicity/immune effector cell-associated neurotoxicity syndrome (ICANS) (Table 2). Rule out other causes of neurologic symptoms. Patients who experience Grade 2 or higher neurologic toxicities/ICANS should be monitored with continuous cardiac telemetry and pulse oximetry.

Provide intensive-care supportive therapy for severe or life-threatening neurologic toxicities. Consider levetiracetam for seizure prophylaxis for any grade of neurologic toxicities.

Table 2. Neurologic Toxicity/ICANS Grading and Management Guidance

Grading Assessment ^a	Concurrent CRS	No Concurrent CRS
Grade 1	<p>Administer tocilizumab per Table 1 for management of Grade 1 CRS.</p> <p>In addition, administer one dose of dexamethasone 10 mg intravenously.</p> <p>If not improving after 2 days, repeat dexamethasone 10 mg intravenously.</p>	<p>Administer one dose of dexamethasone 10 mg intravenously.</p> <p>If not improving after 2 days, repeat dexamethasone 10 mg intravenously.</p>
	<p>Consider levetiracetam for seizure prophylaxis.</p>	
Grade 2	<p>Administer tocilizumab per Table 1 for management of Grade 2 CRS.</p> <p>In addition, administer dexamethasone 10 mg intravenously 4 times a day.</p> <p>If improving, continue corticosteroids until the severity is Grade 1 or less, then quickly taper as clinically appropriate.</p> <p>If not improving, manage as appropriate grade below.</p>	<p>Administer dexamethasone 10 mg intravenously 4 times a day.</p> <p>If improving, continue corticosteroids until the severity is Grade 1 or less, then quickly taper as clinically appropriate.</p> <p>If not improving, manage as appropriate grade below.</p>
	<p>Consider levetiracetam for seizure prophylaxis.</p>	
Grade 3	<p>Administer tocilizumab per Table 1 for management of Grade 2 CRS.</p> <p>In addition, administer methylprednisolone 1000 mg intravenously once daily.</p> <p>If improving, manage as appropriate grade above and continue corticosteroids until the severity is Grade 1 or less, then taper as clinically appropriate.</p> <p>If not improving, manage as Grade 4.</p>	<p>Administer methylprednisolone 1000 mg intravenously once daily.</p> <p>If improving, manage as appropriate grade above and continue corticosteroids until the severity is Grade 1 or less, then taper as clinically appropriate.</p> <p>If not improving, manage as Grade 4.</p>
	<p>Consider levetiracetam for seizure prophylaxis.</p>	

Grading Assessment ^a	Concurrent CRS	No Concurrent CRS
Grade 4	Administer tocilizumab per Table 1 for management of Grade 2 CRS. In addition, administer methylprednisolone 1000 mg intravenously twice per day. If improving, manage as appropriate grade above and continue corticosteroids until the severity is Grade 1 or less, then taper as clinically appropriate. If not improving, consider 1000 mg of methylprednisolone intravenously 3 times a day or alternate therapy. ^b	Administer methylprednisolone 1000 mg intravenously twice per day. If improving, manage as appropriate grade above and continue corticosteroids until the severity is Grade 1 or less, then taper as clinically appropriate. If not improving, consider 1000 mg of methylprednisolone intravenously 3 times a day or alternate therapy. ^b
	Consider levetiracetam for seizure prophylaxis.	

a. Severity based on Common Terminology Criteria for Adverse Events.

b. Alternate therapy includes (but is not limited to): anakinra, siltuximab, ruxolitinib, cyclophosphamide, IVIG and ATG.

3 DOSAGE FORMS AND STRENGTHS

YESCARTA is available as a cell suspension for infusion.

A single dose of YESCARTA contains 2×10^6 CAR-positive viable T cells per kg of body weight (or maximum of 2×10^8 CAR-positive viable T cells for patients 100 kg and above) in approximately 68 mL suspension in an infusion bag [see *How Supplied/Storage and Handling (13)*].

4 CONTRAINDICATIONS

Hypersensitivity to the active substance or to any of the excipients [see *Hypersensitivity Reactions (5.4)*].
Contraindications of the lymphodepleting chemotherapy must be considered.

5 WARNINGS AND PRECAUTIONS

5.1 Cytokine Release Syndrome

CRS, including fatal or life-threatening reactions, occurred following treatment with YESCARTA. CRS occurred in 92% (257/278) of patients with LBCL in ZUMA-7 and ZUMA-1, including \geq Grade 3 (Lee grading system¹) CRS in 8%. [see *Adverse Reactions (6)*]. Among patients with LBCL who died after receiving YESCARTA, four had ongoing CRS events at the time of death. In ZUMA-1, the median time to onset of CRS was 2 days (range: 1 to 12 days) and the median duration was 7 days (range: 2 to 29 days, except for one outlying observation of 58 days). In ZUMA-7, the median time to onset of CRS was 3 days following infusion (range: 1 to 10 days) and the median duration was 7 days (range: 2 to 43 days).

Key manifestations of CRS ($>10\%$) included fever (93%), hypotension (44%), chills (23%), sinus tachycardia (22%), hypoxia (21%), tachycardia (15%) and headache (14%). Serious events that may be associated with CRS include cardiac arrhythmias (including atrial fibrillation and ventricular tachycardia), cardiac arrest, cardiac failure, renal insufficiency, capillary leak syndrome, hypotension, hypoxia and hemophagocytic lymphohistiocytosis/macrophage activation syndrome (HLH/MAS) [see *Adverse Reactions (6)*].

The impact of tocilizumab and/or corticosteroids on the incidence and severity of CRS was assessed in two subsequent cohorts of LBCL patients in ZUMA-1. Among patients who received tocilizumab and/or corticosteroids for ongoing Grade 1 events (see Table 1) [see *Clinical Trials Experience (6.1)*], CRS occurred in 93% (38/41), including 2%

(1/41) with Grade 3 CRS; no patients experienced a Grade 4 or 5 event. The median time to onset of CRS was 2 days (range: 1 to 8 days) and the median duration of CRS was 7 days (range: 2 to 16 days).

Prophylactic treatment with corticosteroids was administered to a cohort of 39 patients for 3 days beginning on the day of infusion of YESCARTA [see *Clinical Trials Experience (6.1)*]. Thirty-one of the 39 patients (79%) developed CRS at which point the patients were managed with tocilizumab and/or therapeutic doses of corticosteroids with no patients developing Grade 3 or higher CRS. The median time to onset of CRS was 5 days (range: 1 to 15 days) and the median duration of CRS was 4 days (range: 1 to 10 days). Although there is no known mechanistic explanation, consider the risk and benefits of prophylactic corticosteroids in the context of pre-existing comorbidities for the individual patient and the potential for the risk of Grade 4 and prolonged neurologic toxicities [See *Neurologic Toxicities (5.2)*].

Ensure that 2 doses of tocilizumab are available prior to infusion of YESCARTA. Monitor patients at least daily for 7 days at the certified healthcare facility following infusion for signs and symptoms of CRS. Monitor patients for signs or symptoms of CRS for 4 weeks after infusion. Counsel patients to seek immediate medical attention should signs or symptoms of CRS occur at any time [see *Patient Counseling Information (14)*]. At the first sign of CRS, institute treatment with supportive care, tocilizumab, or tocilizumab and corticosteroids as indicated [see *Dosage and Administration (2.3)*].

5.2 Neurologic Toxicities

Neurologic toxicities (including ICANS) that were fatal or life-threatening occurred following treatment with YESCARTA.

Neurologic toxicities occurred in 62% (173/278) of patients with LBCL, including \geq Grade 3 cases in 25%. The median time to onset for neurologic toxicity was 5 days (range: 1 to 17 days) and the median duration was 13 days in patients in ZUMA-1. The median time to onset for neurologic toxicity was 5 days (range: 1 to 133 days) and median duration was 14 days in patients in ZUMA-7. Neurologic toxicities occurred in 98% of patients with LBCL within the first 8 weeks of YESCARTA infusion and in 87% within the first 7 days. Neurologic events resolved in all but 4 subjects who had ongoing neurologic events at the time of death.

The most common neurologic toxicities ($>10\%$) included tremor (28%), confusional state (25%), encephalopathy (24%), aphasia (20%), and somnolence (13%). Prolonged encephalopathy lasting up to 173 days was noted. Serious events including leukoencephalopathy and seizures occurred with YESCARTA. Fatal and serious cases of cerebral edema and encephalopathy, including late-onset encephalopathy, have occurred in patients treated with YESCARTA.

The impact of tocilizumab and/or corticosteroids on the incidence and severity of neurologic toxicities was assessed in two subsequent cohorts of LBCL patients in ZUMA-1. Among patients who received corticosteroids at the onset of Grade 1 toxicities (see Table 2), neurologic toxicities occurred in 78% (32/41) and 20% (8/41) had Grade 3 neurologic toxicities; no patients experienced a Grade 4 or 5 event. The median time to onset of neurologic toxicities was 6 days (range: 1 to 93 days) with a median duration of 8 days (range: 1 to 144 days).

Prophylactic treatment with corticosteroids was administered to a cohort of 39 patients for 3 days beginning on the day of infusion of YESCARTA [see *Clinical Trials Experience (6.1)*]. Of these 39 patients, 85% (33/39) developed neurologic toxicities, 8% (3/39) developed Grade 3 and 5% (2/39) developed Grade 4 neurologic toxicities. The median time to onset of neurological toxicities was 6 days (range: 1 to 274 days) with a median duration of 12 days (range: 1 to 107 days). Prophylactic corticosteroids for management of CRS and neurologic toxicities may result in higher grade of neurologic toxicities or prolongation of neurologic toxicities, delay the onset and decrease the duration of CRS [See *Cytokine Release Syndrome (5.1)*].

Monitor patients at least daily for 7 days at the certified healthcare facility following infusion for signs and symptoms of neurologic toxicities. Monitor patients for signs or symptoms of neurologic toxicities for 4 weeks after infusion and treat promptly [see *Dosage and Administration (2.3)*].

5.3 YESCARTA Controlled Distribution Program

Because of the risk of CRS and neurologic toxicities, YESCARTA is available only through a Controlled Distribution Program [see *Boxed Warning and Warnings and Precautions (5.1 and 5.2)*]. The required components of the YESCARTA Controlled Distribution Program are:

- Healthcare facilities that dispense and administer YESCARTA must be enrolled and comply with the Controlled Distribution Program requirements. Certified healthcare facilities must have on-site, immediate access to tocilizumab, and ensure that a minimum of 2 doses of tocilizumab are available for each patient for infusion within 2 hours after YESCARTA infusion, if needed for treatment of CRS.
- Certified healthcare facilities must ensure that healthcare providers who prescribe, dispense, or administer YESCARTA are trained about the management of CRS and neurologic toxicities.

5.4 Hypersensitivity Reactions

Allergic reactions may occur with the infusion of YESCARTA. Serious hypersensitivity reactions, including anaphylaxis, may be due to dimethyl sulfoxide (DMSO) or residual gentamicin in YESCARTA.

5.5 Serious Infections

Severe or life-threatening infections occurred in patients after YESCARTA infusion. Infections (all grades) occurred in 41% of LBCL patients treated. Grade 3 or higher infections occurred in 19% of patients, including Grade 3 or higher infections with an unspecified pathogen in 13%, bacterial infections in 6%, viral infections in 5% and opportunistic infections in 1%. YESCARTA should not be administered to patients with clinically significant active systemic infections. Monitor patients for signs and symptoms of infection before and after YESCARTA infusion and treat appropriately. Administer prophylactic antimicrobials according to local guidelines.

Febrile neutropenia was observed in 15% of patients after YESCARTA infusion and may be concurrent with CRS. In the event of febrile neutropenia, evaluate for infection and manage with broad-spectrum antibiotics, fluids, and other supportive care as medically indicated.

In immunosuppressed patients, including those who have received YESCARTA, life-threatening and fatal opportunistic infections including disseminated fungal infections (e.g., candida sepsis and aspergillus infections) and viral reactivation (e.g., human herpes virus-6 [HHV-6] encephalitis and JC virus progressive multifocal leukoencephalopathy [PML]) have been reported. The possibility of HHV-6 encephalitis and PML should be considered in immunosuppressed patients with neurologic events and appropriate diagnostic evaluations should be performed.

Hepatitis B Virus Reactivation

Hepatitis B virus (HBV) reactivation, in some cases resulting in fulminant hepatitis, hepatic failure, and death, has occurred in patients treated with drugs directed against B cells, including YESCARTA. Perform screening for HBV, HCV, and HIV and management in accordance with clinical guidelines before collection of cells for manufacturing.

5.6 Prolonged Cytopenias

Patients may exhibit cytopenias for several weeks following lymphodepleting chemotherapy and YESCARTA infusion. Grade 3 or higher cytopenias not resolved by Day 30 following YESCARTA infusion occurred in 33% of patients and included neutropenia (26%), thrombocytopenia (13%), and anemia (6%). Monitor blood counts after YESCARTA infusion.

5.7 Hypogammaglobulinemia

B-cell aplasia and hypogammaglobulinemia can occur in patients receiving treatment with YESCARTA. Hypogammaglobulinemia was reported as an adverse reaction in 13% of patients. Monitor immunoglobulin levels after treatment with YESCARTA and manage using infection precautions, antibiotic prophylaxis, and immunoglobulin replacement.

The safety of immunization with live viral vaccines during or following YESCARTA treatment has not been studied. Vaccination with live virus vaccines is not recommended for at least 6 weeks prior to the start of lymphodepleting chemotherapy, during YESCARTA treatment, and until immune recovery following treatment with YESCARTA.

5.8 Secondary Malignancies

Patients treated with YESCARTA may develop secondary malignancies. Monitor life-long for secondary malignancies. In the event that a secondary malignancy of T-cell origin occurs, contact Kite at asiamedinfo@gilead.com to obtain instructions on patient samples to collect for testing.

5.9 Tumour Lysis Syndrome (TLS)

TLS, which may be severe, has occasionally been observed. To minimise risk of TLS, patients with elevated uric acid or high tumour burden should receive allopurinol, or an alternative prophylaxis, prior to YESCARTA infusion. Signs and symptoms of TLS should be monitored, and events managed according to standard guidelines.

5.10 Graft versus Host Disease (GvHD)

Due to the risks associated with YESCARTA treatment, infusion must be delayed if a patient has active or chronic GvHD following allogeneic stem cell transplant.

5.11 Effects on Ability to Drive and Use Machines

Due to the potential for neurologic events, including altered mental status or seizures, patients receiving YESCARTA are at risk for altered or decreased consciousness or coordination in the 8 weeks following YESCARTA infusion. Advise patients to refrain from driving and engaging in hazardous occupations or activities, such as operating heavy or potentially dangerous machinery, during this initial period.

6 ADVERSE REACTIONS

The following adverse reactions are described elsewhere in the labeling:

- Cytokine Release Syndrome [*see Warnings and Precautions (5.1, 5.3)*]
- Neurologic Toxicities [*see Warnings and Precautions (5.2, 5.3)*]
- Hypersensitivity Reactions [*see Warnings and Precautions (5.4)*]
- Serious Infections [*see Warnings and Precautions (5.5)*]
- Prolonged Cytopenias [*see Warnings and Precautions (5.6)*]
- Hypogammaglobulinemia [*see Warnings and Precautions (5.7)*]

6.1 Clinical Trials Experience

Because clinical trials are conducted under widely varying conditions, adverse reaction rates observed in the clinical trials of a drug cannot be directly compared to rates in the clinical trials of another drug and may not reflect the rates observed in practice.

The data described in the WARNINGS AND PRECAUTIONS reflect exposure to a single dose of YESCARTA in one randomized, open-label study with 168 patients with relapsed or refractory LBCL (ZUMA-7) and one open-label, single-arm study in which 108 patients with relapsed or refractory LBCL (ZUMA-1).

Relapsed or Refractory Large B-cell Lymphoma

ZUMA-7

The safety of YESCARTA was evaluated in ZUMA-7, a randomized, open-label, multicenter study in which patients with primary refractory LBCL or first relapse of LBCL received YESCARTA (N = 168) or standard therapy (N = 168) [see *Clinical Studies (14)*]. Patients had not yet received treatment for relapsed or refractory lymphoma and were potential candidates for autologous HSCT. The trial excluded patients who were not deemed candidates for transplant or who had a history of central nervous system (CNS) disorders (such as seizures or cerebrovascular ischemia), serious or uncontrolled infection, or autoimmune disease requiring systemic immunosuppression. The study required ANC \geq 1000/mm³, platelet count \geq 75,000/mm³, creatinine clearance \geq 60 ml/min, AST/ALT \leq 2.5 x ULN, and total bilirubin \leq 1.5mg/dL.

The median age of the YESCARTA-treated safety population was 59 years (range: 21 to 80 years); 62% were male. The baseline Eastern Cooperative Oncology Group (ECOG) performance status was 0 in 54% of patients and 1 in 46%.

The most common non-laboratory adverse reactions to YESCARTA (incidence \geq 20%) included fever, CRS, fatigue, hypotension, encephalopathy, tachycardia, diarrhea, headache, musculoskeletal pain, nausea, febrile neutropenia, chills, cough, infection with unspecified pathogen, dizziness, tremor, decreased appetite, edema, hypoxia, abdominal pain, aphasia, constipation, and vomiting. Serious adverse reactions occurred in 50% of patients. The most common serious adverse reactions ($>$ 5%) included CRS, fever, encephalopathy, hypotension, infection with unspecified pathogen, and pneumonia. Fatal adverse reactions occurred in 2% of patients.

The most common (\geq 10%) Grade 3 or higher non-laboratory adverse reactions included febrile neutropenia, encephalopathy, and hypotension.

Sixty-seven percent (112/168) of patients received tocilizumab after infusion of YESCARTA.

Table 3 summarizes selected non-laboratory adverse reactions in patients treated with YESCARTA, and Table 4 summarizes selected new or worsening Grade 3 or 4 laboratory abnormalities.

Table 3. Adverse Reactions in \geq 10% of Patients Treated with YESCARTA in ZUMA-7

Adverse Reaction	YESCARTA N = 168	
	Any Grade (%)	Grade 3 or Higher (%)
Febrile neutropenia	31	31
<i>Cardiac Disorders</i>		
Tachycardia ^a	43	2
Arrhythmia ^b	14	3
<i>Gastrointestinal Disorders</i>		
Diarrhea ^c	42	3
Nausea	40	2
Abdominal pain ^d	20	4
Constipation	20	0
Vomiting	20	0
Dry Mouth	10	0
<i>General Disorders and Administration Site Conditions</i>		
Fever ^e	93	9
Fatigue ^f	52	7
Chills	28	1
Edema ^g	23	1
<i>Immune System Disorders</i>		
Cytokine release syndrome	92	7

Adverse Reaction	YESCARTA N = 168	
	Any Grade (%)	Grade 3 or Higher (%)
Hypogammaglobulinemia	11	0
<i>Infections and Infestations</i>		
Infections with pathogen unspecified	25	8
Viral infections	15	4
Bacterial infections	10	5
Fungal infections	10	1
<i>Metabolism and Nutrition Disorders</i>		
Decreased appetite	24	4
<i>Musculoskeletal and Connective Tissue Disorders</i>		
Musculoskeletal pain ^h	40	1
Motor dysfunction ⁱ	15	4
<i>Nervous System Disorders</i>		
Encephalopathy ^j	46	18
Headache ^k	41	3
Tremor	25	1
Dizziness ^l	25	4
Aphasia	20	7
Neuropathy peripheral ^m	11	2
<i>Psychiatric Disorders</i>		
Insomnia ⁿ	13	0
Delirium ^o	12	4
<i>Renal and Urinary Disorders</i>		
Renal insufficiency ^p	11	2
<i>Respiratory, Thoracic and Mediastinal Disorders</i>		
Cough ^q	27	1
Hypoxia	21	9
<i>Skin and Subcutaneous Tissue Disorders</i>		
Rash ^r	17	1
<i>Vascular Disorders</i>		
Hypotension ^s	47	11

The following events were also counted in the incidence of CRS: coagulopathy, tachycardia, arrhythmia, cardiac failure, diarrhea, nausea, vomiting, fever, fatigue, chills, edema, decreased appetite, musculoskeletal pain, headache, tremor, dizziness, renal insufficiency, cough, hypoxia, dyspnea, pleural effusion, respiratory failure, rash, hypotension, and hypertension.

- a. Tachycardia includes tachycardia, sinus tachycardia.
- b. Arrhythmia includes arrhythmia, atrial fibrillation, bradycardia, electrocardiogram QT prolonged, extrasystoles, sinus bradycardia, supraventricular extrasystoles, supraventricular tachycardia, ventricular extrasystoles, ventricular tachycardia.
- c. Diarrhea includes diarrhea, colitis.
- d. Abdominal pain includes abdominal pain, abdominal discomfort, abdominal pain lower, abdominal pain upper, dyspepsia.
- e. Fever includes pyrexia.
- f. Fatigue includes fatigue, asthenia, malaise.
- g. Edema includes edema, face edema, fluid overload, generalized edema, hypervolemia, localized edema, edema genital, edema peripheral, periorbital edema, peripheral swelling, pulmonary edema.
- h. Musculoskeletal pain includes musculoskeletal pain, arthralgia, arthritis, back pain, bone pain, flank pain, groin pain, musculoskeletal chest pain, myalgia, neck pain, non-cardiac chest pain, pain in extremity.
- i. Motor dysfunction includes muscle contractions involuntary, muscle spasms, muscle twitching, muscular weakness.
- j. Encephalopathy includes encephalopathy, altered state of consciousness, amnesia, apraxia, bradyphrenia, cognitive disorder, confusional state, depressed level of consciousness, disturbance in attention, dysarthria, dysgraphia, dyspraxia, lethargy, loss of consciousness, memory impairment, mental impairment, mental status changes, metabolic encephalopathy, slow speech, somnolence, toxic encephalopathy.
- k. Headache includes headache and tension headache.
- l. Dizziness includes dizziness, dizziness postural, presyncope, syncope, vertigo.

- m. Neuropathy peripheral includes hypoesthesia, lumbar radiculopathy, neuropathy peripheral, paresthesia, peroneal nerve palsy, sciatica.
- n. Insomnia includes insomnia and sleep deficit.
- o. Delirium includes delirium, agitation, delusion, disorientation, hallucination, irritability, restlessness.
- p. Renal insufficiency includes acute kidney injury, blood creatinine increased, chronic kidney disease.
- q. Cough includes cough, productive cough, upper-airway cough syndrome.
- r. Rash includes rash, dermatitis, dermatitis allergic, dermatitis bullous, drug eruption, erythema, pruritus, rash macular, rash maculo-papular, rash pruritic, urticaria.
- s. Hypotension includes hypotension, capillary leak syndrome, orthostatic hypotension.

Other clinically important adverse reactions that occurred in less than 10% of patients treated with YESCARTA include the following:

- *Blood and lymphatic system disorders:* Coagulopathy (9%)
- *Cardiac disorders:* Cardiac failure (1%)
- *Eye Disorders:* Visual impairment (7%)
- *Infections and infestations:* Pneumonia (8%), Sepsis (4%)
- *Nervous system disorders:* Ataxia (6%), seizure (3%), myoclonus (2%), facial paralysis (2%), paresis (2%)
- *Respiratory, thoracic and mediastinal disorders:* Dyspnea (8%), pleural effusion (6%), respiratory failure (2%)
- *Vascular disorders:* Hypertension (9%), thrombosis (7%)

Laboratory abnormalities:

Table 4. Grade 3 or 4 Laboratory Abnormalities Occurring in $\geq 10\%$ of Patients in ZUMA-7 Following Treatment with YESCARTA^a (N = 168)

Laboratory Abnormality	YESCARTA
	Grades 3 or 4 (%)
Leukocyte decrease	95
Neutrophil decrease	94
Lymphocyte decrease	94
Hemoglobin decrease	40
Platelet decrease	26
Sodium decrease	12
Glucose increase	11

a. Baseline lab values were assessed prior to lymphodepleting chemotherapy.

ZUMA-1

The safety of YESCARTA was evaluated in ZUMA-1, a study in which 108 patients with relapsed or refractory LBCL received CD19-positive CAR T cells based on a recommended dose which was weight-based [see *Clinical Studies (11)*]. Patients with a history of CNS disorders (such as seizures or cerebrovascular ischemia) or autoimmune disease requiring systemic immunosuppression were ineligible. The median age of the study population was 58 years (range: 23 to 76 years); 68% were male. The baseline Eastern Cooperative Oncology Group (ECOG) performance status was 0 in 43% of patients and 1 in 57% of patients.

The most common adverse reactions (incidence $\geq 20\%$) included CRS, encephalopathy, fatigue, decreased appetite, headache, fever, febrile neutropenia, diarrhea, nausea, tremor, tachycardia, cough, other pathogen infections, hypotension, vomiting, viral infections, dizziness, constipation, and edema. Serious adverse reactions occurred in 49% of patients. The most common serious adverse reactions ($> 2\%$) included encephalopathy, other pathogen infections, CRS, bacterial infections, fever, viral infections, aphasia, delirium, cardiac arrest, and dyspnea.

The most common ($\geq 10\%$) Grade 3 or higher reactions included febrile neutropenia, encephalopathy, other pathogen infections, and CRS.

Forty-five percent (49/108) of patients received tocilizumab after infusion of YESCARTA.

Table 5 summarizes non-laboratory adverse reactions that occurred in $\geq 10\%$ of patients treated with YESCARTA, and Table 6 describes the laboratory abnormalities of Grade 3 or 4 that occurred in $\geq 10\%$ of patients.

Table 5. Summary of Adverse Reactions Observed in $\geq 10\%$ of Patients Treated with YESCARTA in ZUMA-1 (N = 108)

Adverse Reaction	Any Grade (%)	Grade 3 or Higher (%)
<i>Blood Disorders</i>		
Febrile neutropenia	36	32
<i>Cardiac Disorders</i>		
Tachycardia ^a	29	1
Arrhythmia ^b	18	3
<i>Gastrointestinal Disorders</i>		
Diarrhea	35	4
Nausea	31	0
Vomiting	24	1
Constipation	20	0
Abdominal pain ^c	15	2
Dry mouth	11	0
<i>General Disorders and Administration Site Conditions</i>		
Fatigue ^d	43	3
Fever	39	3
Edema ^e	20	1
Chills	19	0
<i>Immune System Disorders</i>		
Cytokine release syndrome	93	11
Hypogammaglobulinemia ^f	16	0
<i>Infections and Infestations</i>		
Other pathogen infections	28	19
Viral infections	21	6
Bacterial infections	15	9
<i>Investigations</i>		
Decreased appetite	41	2
Weight decreased	15	0
Dehydration	11	3
<i>Musculoskeletal and Connective Tissue Disorders</i>		
Motor dysfunction ^g	17	1
Pain in extremity ^h	17	1
Back pain	14	1
Muscle pain	10	1
Arthralgia	10	0
<i>Nervous System Disorders</i>		
Encephalopathy ⁱ	58	31
Headache ^j	40	1
Tremor	31	2
Dizziness ^k	21	0
Aphasia	18	7
<i>Psychiatric Disorders</i>		

Adverse Reaction	Any Grade (%)	Grade 3 or Higher (%)
Delirium ^l	17	6
Anxiety	11	1
<i>Respiratory, Thoracic and Mediastinal Disorders</i>		
Cough ^m	29	0
Dyspnea ⁿ	17	3
Hypoxia ^o	14	2
Pleural effusion	13	2
<i>Vascular Disorders</i>		
Hypotension ^p	27	6
Hypertension	15	6

- a. Tachycardia includes tachycardia, sinus tachycardia.
- b. Arrhythmia includes arrhythmia, atrial fibrillation, atrial flutter, atrioventricular block, bundle branch block right, electrocardiogram QT prolonged, extra-systoles, heart rate irregular, supraventricular extra systoles, supraventricular tachycardia, ventricular arrhythmia, ventricular tachycardia.
- c. Abdominal pain includes abdominal pain, abdominal discomfort, abdominal pain lower, abdominal pain upper, abdominal tenderness.
- d. Fatigue includes fatigue, malaise.
- e. Edema includes face edema, generalized edema, swelling, localized edema, edema, edema genital, edema peripheral, periorbital edema, peripheral swelling, scrotal edema.
- f. Hypogammaglobulinemia includes hypogammaglobulinemia, blood immunoglobulin G decreased.
- g. Motor dysfunction includes muscle spasms, muscular weakness.
- h. Pain in extremity includes pain, pain in extremity.
- i. Encephalopathy includes cognitive disorder, confusional state, depressed level of consciousness, disturbance in attention, encephalopathy, hypersomnia, leukoencephalopathy, memory impairment, mental status changes, paranoia, somnolence, stupor, toxic encephalopathy.
- j. Headache includes headache, head discomfort, procedural headache.
- k. Dizziness includes dizziness, presyncope, syncope.
- l. Delirium includes agitation, delirium, delusion, disorientation, hallucination, irritability, restlessness.
- m. Cough includes cough, productive cough, upper-airway cough syndrome.
- n. Dyspnea includes acute respiratory failure, dyspnea, orthopnea, respiratory distress.
- o. Hypoxia includes hypoxia, oxygen saturation decreased.
- p. Hypotension includes diastolic hypotension, hypotension, orthostatic hypotension.

Other clinically important adverse reactions that occurred in less than 10% of patients treated with YESCARTA include the following:

- *Blood disorders:* Coagulopathy (2%)
- *Cardiac disorders:* Cardiac failure (2%), cardiac arrest (3%)
- *Immune system disorders:* Hemophagocytic lymphohistiocytosis/macrophage activation syndrome (HLH/MAS) (1%), hypersensitivity (2%)
- *Infections and infestations disorders:* Fungal infections (6%)
- *Nervous system disorders:* Ataxia (6%), neuropathy (6%), seizure (4%), dyscalculia (2%), myoclonus (2%)
- *Respiratory, thoracic and mediastinal disorders:* Pulmonary edema (7%)
- *Renal and urinary disorders:* Renal insufficiency (8%)
- *Skin and subcutaneous tissue disorders:* Rash (6%)
- *Vascular disorders:* Thrombosis (6%), Capillary leak syndrome (1%)

Laboratory abnormalities:

Table 6. Grade 3 or 4 Laboratory Abnormalities Occurring in $\geq 10\%$ of Patients in ZUMA-1 Following Treatment with YESCARTA^a (N = 108)

Laboratory Abnormality	Grades 3 or 4 (%)
Lymphocyte decrease	99
Leukocyte decrease	96
Neutrophil decrease	94
Hemoglobin decrease	65
Platelet decrease	56
Phosphate decrease	52
Sodium decrease	23
Uric acid increased	15
Direct bilirubin increased	13
Alanine aminotransferase increased	12
Potassium decrease	11
Aspartate Aminotransferase increased	10

a. Baseline lab values were assessed prior to lymphodepleting chemotherapy.

The safety and efficacy of YESCARTA was evaluated in two subsequent cohorts of LBCL patients. The first subsequent, open label, safety management cohort in ZUMA-1 evaluated the safety and efficacy of YESCARTA with the use of tocilizumab and/or corticosteroid and prophylactic levetiracetam (750mg PO or IV twice daily) for Grade 1 CRS or neurologic events (see Tables 1 and 2). A total of 46 patients with relapsed or refractory LBCL were enrolled and 41 patients were treated with YESCARTA. Of the remaining 5 patients who were not treated, 2 patients died prior to receiving YESCARTA and 3 patients were ineligible due to disease progression. Twenty-eight patients (68%) treated with YESCARTA received bridging therapy between leukapheresis and lymphodepleting chemotherapy. Thirty-two patients (78%) treated with YESCARTA received tocilizumab and /or corticosteroid for CRS and/or neurologic events. Fifteen of 36 with Grade 1 CRS and 21 of 24 patients with Grade 2 CRS received tocilizumab and/or corticosteroids. Among patients who received treatment for Grade 1 or Grade 2 CRS, most patients (13 of 15 and 19 of 21 patients, respectively) received both tocilizumab and corticosteroids. Most patients received 1 or 2 doses of each drug. Ten of 27 patients with Grade 1 and 7 of 15 patients with Grade 2 neurologic events received corticosteroids alone or in combination with tocilizumab.

The second subsequent, open label, safety management cohort in ZUMA-1 evaluated the safety and efficacy of YESCARTA with the use of prophylactic corticosteroids (oral dexamethasone 10 mg once daily for 3 days, starting prior to YESCARTA infusion on Day 0) and prophylactic levetiracetam (750 mg PO or IV) [*see Warnings and Precautions (5.1 and 5.2)*].

6.2 Immunogenicity

YESCARTA has the potential to induce anti-product antibodies. The immunogenicity of YESCARTA has been evaluated using an enzyme-linked immunosorbent assay (ELISA) for the detection of binding antibodies against FMC63, the originating antibody of the anti-CD19 CAR. Eleven patients (4%) tested positive for pre-dose anti-FMC63 antibodies at baseline in ZUMA-7 and ZUMA-1, and one patient (1%) who had a negative test result at baseline had a positive test result post administration of YESCARTA in the screening ELISA in ZUMA-7. There is no evidence that the kinetics of initial expansion and persistence of YESCARTA, or the safety or effectiveness of YESCARTA, was altered in these patients.

6.3 Postmarketing Experience

The following adverse reactions have been identified during post-approval use of YESCARTA. Because these reactions are reported voluntarily from a population of uncertain size, it is not always possible to reliably estimate their frequency or establish a causal relationship to drug exposure.

Nervous System Disorders

Spinal cord edema, myelitis, quadriplegia, dysphagia, ICANS, and status epilepticus.

Immune System Disorders

Infusion related reaction.

7 USE IN SPECIFIC POPULATIONS

7.1 Pregnancy

Risk Summary

There are no available data with YESCARTA use in pregnant women. No animal reproductive and developmental toxicity studies have been conducted with YESCARTA to assess whether it can cause fetal harm when administered to a pregnant woman. It is not known if YESCARTA has the potential to be transferred to the fetus. Based on the mechanism of action, if the transduced cells cross the placenta, they may cause fetal toxicity, including B-cell lymphocytopenia. Therefore, YESCARTA is not recommended for women who are pregnant, and pregnancy after YESCARTA infusion should be discussed with the treating physician.

7.2 Lactation

Risk Summary

There is no information regarding the presence of YESCARTA in human milk, the effect on the breastfed infant, and the effects on milk production. The developmental and health benefits of breastfeeding should be considered along with the mother's clinical need for YESCARTA and any potential adverse effects on the breastfed infant from YESCARTA or from the underlying maternal condition.

7.3 Females and Males of Reproductive Potential

Pregnancy Testing

Pregnancy status of females with reproductive potential should be verified. Sexually active females of reproductive potential should have a pregnancy test prior to starting treatment with YESCARTA.

Contraception

See the package insert for fludarabine and cyclophosphamide for information on the need for effective contraception in patients who receive the lymphodepleting chemotherapy.

There are insufficient exposure data to provide a recommendation concerning duration of contraception following treatment with YESCARTA.

Infertility

There are no data on the effect of YESCARTA on fertility.

7.4 Pediatric Use

The safety and efficacy of YESCARTA have not been established in pediatric patients.

7.5 Geriatric Use

No clinically important differences in safety or effectiveness were observed between patients aged 65 years and older and younger patients.

8 DESCRIPTION

YESCARTA (axicabtagene ciloleucel) is a CD19-directed genetically modified autologous T cell immunotherapy. To prepare YESCARTA, a patient's own T cells are harvested and genetically modified *ex vivo* by retroviral transduction to express a chimeric antigen receptor (CAR) comprising a murine anti-CD19 single chain variable fragment (scFv) linked to CD28 and CD3-zeta co-stimulatory domains. The anti-CD19 CAR T cells are expanded and infused back into the patient, where they can recognize and eliminate CD19-expressing target cells.

YESCARTA is prepared from the patient's peripheral blood mononuclear cells, which are obtained via a standard leukapheresis procedure. The mononuclear cells are enriched for T cells and activated with anti-CD3 antibody in the presence of IL-2, then transduced with the replication incompetent retroviral vector containing the anti-CD19 CAR transgene. The transduced T cells are expanded in cell culture, washed, formulated into a suspension, and cryopreserved. The product must pass a sterility test before release for shipping as a frozen suspension in a patient-specific infusion bag. The product is thawed prior to infusion [*see Dosage and Administration (2.2), How Supplied/Storage and Handling (13)*].

In addition to T cells, YESCARTA may contain NK and NK-T cells. Cryostor CS10 (contains DMSO), sodium chloride and human albumin are present as excipients.

YESCARTA is a clear to opaque, white to red dispersion for infusion, supplied in an infusion bag individually packed in a metal cassette.

9 CLINICAL PHARMACOLOGY

9.1 Mechanism of Action

YESCARTA, a CD19-directed genetically modified autologous T cell immunotherapy, binds to CD19-expressing cancer cells and normal B cells. Studies demonstrated that following anti-CD19 CAR T cell engagement with CD19-expressing target cells, the CD28 and CD3-zeta co-stimulatory domains activate downstream signaling cascades that lead to T cell activation, proliferation, acquisition of effector functions and secretion of inflammatory cytokines and chemokines. This sequence of events leads to killing of CD19-expressing cells.

9.2 Pharmacodynamics

After YESCARTA infusion, pharmacodynamic responses were evaluated over a 4-week interval by measuring transient elevation of cytokines, chemokines and other molecules in blood. Levels of cytokines and chemokines such as IL-6, IL-8, IL-10, IL-15, TNF- α , IFN- γ , and sIL2R α were analyzed. Peak elevation was observed within the first 14 days after infusion, and levels generally returned to baseline within 28 days.

Due to the on-target effect of YESCARTA, a period of B-cell aplasia is expected.

Large B-cell lymphoma

Among patients with LBCL with an ongoing response at 24 months in the ZUMA-7 study, 21 of 61 evaluable patients (34%) had no detectable B cells at baseline, and the majority of patients at Month 3 (43 of 69 evaluable patients [62%]) and Month 6 (8 of 13 evaluable patients [62%]) had no detectable B cells. At Month 24, 20 of 24 evaluable patients (83%) had detectable B cells.

Among patients with LBCL with an ongoing response at 24 months in the ZUMA-1 study, 13 of 29 evaluable patients (45%) had no detectable B cells at baseline, and the majority of patients at Month 3 (28 of 35 evaluable patients [80%]) and Month 6 (25 of 32 evaluable patients [78%]) had no detectable B cells. At Month 24, 24 of 32 evaluable patients (75%) had detectable B cells.

9.3 Pharmacokinetics

Following infusion of YESCARTA, anti-CD19 CAR T cells exhibited an initial rapid expansion followed by a decline to near baseline levels by 3 months. Peak levels of anti-CD19 CAR T cells occurred within the first 7 - 14 days after YESCARTA infusion.

Age (range: 21 to 80 years) and gender had no significant impact on AUC Day 0 - 28 and C_{max} of YESCARTA.

Large B-cell Lymphoma

Among patients with LBCL in the ZUMA-7 study (n=162 evaluable), the number of anti-CD19 CAR T cells in blood was positively associated with objective response [complete remission (CR) or partial remission (PR)]. The median anti-CD19 CAR T cell C_{max} levels in responders (n=142) were 275% higher compared to the corresponding level in nonresponders (n=20) (28.9 cells/ μ L vs 10.5 cells/ μ L). Median AUC Day 0 - 28 in responding patients (n=142) was 418% of the corresponding level in nonresponders (n=20) (292.9 days \times cells/ μ L vs. 70.1 days \times cells/ μ L).

Among patients with LBCL in the ZUMA-1 study (n=96 evaluable), the number of anti-CD19 CAR T cells in blood was positively associated with objective response (CR or PR). The median anti-CD19 CAR T cell C_{max} levels in responders (n=73) were 205% higher compared to the corresponding level in non-responders (n=23) (43.6 cells/ μ L vs 21.2 cells/ μ L). Median AUC Day 0 - 28 in responding patients (n=73) was 251% of the corresponding level in non-responders (n=23) (557.1 days \times cells/ μ L vs. 222.0 days \times cells/ μ L). No association between peak anti-CD19 CAR T-cell levels and OS was observed among 163 subjects with an evaluable pharmacokinetic sample when anti-CD19 CAR T-cell peaks were categorized as $>$ median relative to \leq median.

Some patients required tocilizumab and corticosteroids for management of CRS and neurologic toxicities. Patients treated with tocilizumab (n=44) had 262% and 232% higher anti-CD19 CAR T cells as measured by AUC Day 0 - 28 and C_{max} respectively, as compared to patients who did not receive tocilizumab (n=57). Similarly, patients that received corticosteroids (n=26) had 217% and 155% higher AUC Day 0 - 28 and C_{max} compared to patients who did not receive corticosteroids (n=75).

Hepatic and renal impairment studies of YESCARTA were not conducted.

10 NONCLINICAL TOXICOLOGY

10.1 Carcinogenesis, Mutagenesis, Impairment of Fertility

No carcinogenicity or genotoxicity studies have been conducted with YESCARTA. No studies have been conducted to evaluate the effects of YESCARTA on fertility.

11 CLINICAL STUDIES

11.1 Relapsed or Refractory Large B-Cell Lymphoma

ZUMA-7

A randomized, open-label, multicenter trial evaluated the efficacy of YESCARTA in adult patients with relapsed or refractory LBCL after first-line chemoimmunotherapy that included rituximab and anthracycline (ZUMA-7; NCT03391466). Patients had not yet received treatment for relapsed or refractory lymphoma and were potential candidates for autologous HSCT. Patients were required to have primary refractory disease or relapse within 12 months following completion of first-line therapy. The study excluded patients with primary mediastinal B-cell lymphoma, any history of central nervous system lymphoma, need for urgent therapy due to tumor mass effect, active or serious infections, and ECOG performance status of 2 or greater.

In total, 359 patients were randomized in a 1:1 ratio to receive a single infusion of YESCARTA or to receive second-line standard therapy, consisting of 2 or 3 cycles of chemoimmunotherapy followed by high-dose therapy and

autologous HSCT in patients who attained CR or PR. Randomization was stratified by response to first-line therapy and second-line age-adjusted International Prognostic Index.

Following lymphodepleting chemotherapy, YESCARTA was administered as a single intravenous infusion at a target dose of 2×10^6 CAR-positive viable T cells/kg (maximum permitted dose: 2×10^8 cells). The lymphodepleting regimen consisted of cyclophosphamide 500 mg/m² intravenously and fludarabine 30 mg/m² intravenously, both given on the fifth, fourth, and third day before YESCARTA. All patients who received YESCARTA were monitored at a healthcare facility for a minimum of 7 days. Bridging therapy, administered between leukapheresis and lymphodepleting chemotherapy, was limited to corticosteroids and was permitted for patients with high disease burden.

In the overall study population, the median age was 59 years (range: 21 to 81 years), 66% were male, 83% were white, 6% were Asian, and 5% were Black. The diagnoses included de novo DLBCL NOS (63%), HGBL with or without *MYC* and *BCL-2* and/or *BCL-6* rearrangements (19%), and large cell transformation of follicular lymphoma (13%). In total, 74% of patients had primary refractory LBCL, and 26% had relapsed disease within 12 months of first-line therapy.

Of the 180 patients randomized to receive YESCARTA, 178 underwent leukapheresis and 170 were treated with YESCARTA, of whom 60 (33%) received bridging corticosteroid therapy. Eight patients (4%) were not treated following leukapheresis, primarily due to progressive disease, serious adverse events, or death. The median time from leukapheresis to product delivery was 18 days (range: 13 to 49 days), and from leukapheresis to YESCARTA infusion was 26 days (range: 16 to 52 days). The median dose was 2.0×10^6 CAR-positive viable T cells/kg (range: 1.0 to 2.1×10^6 cells/kg).

Of the 179 patients randomized to receive standard therapy, 168 patients received any study treatment, and 62 (35%) received high-dose therapy and on-protocol HSCT. The most common reason for not receiving HSCT was lack of response to salvage chemotherapy.

The primary efficacy measure was event-free survival (EFS) as determined by an independent review committee. Efficacy is summarized in Table 7 and Figure 1. The 24-month EFS was 40.5% [95% CI: 33.2, 47.7] in the YESCARTA arm and 16.3% [95% CI: 11.1, 22.2] in the standard therapy arm.

In the YESCARTA arm, the estimated median DOR was 28.4 months (95% CI: 26.9, NE) in patients who achieved CR and 1.6 months (95% CI: 1.4, 1.9) in patients who achieved a best response of PR.

At the time of the primary EFS analysis, the interim analysis of overall survival (OS) did not meet the criteria for statistical significance. With an estimated median follow-up of 46.7 months overall, the primary analysis of OS showed a statistically significant improvement in the YESCARTA arm compared to the standard therapy arm. Fifty-seven percent of patients received cellular immunotherapy after no response to or relapse after randomization to standard therapy arm.

The efficacy results are summarized on Table 7 and Figure 1 and Figure 2.

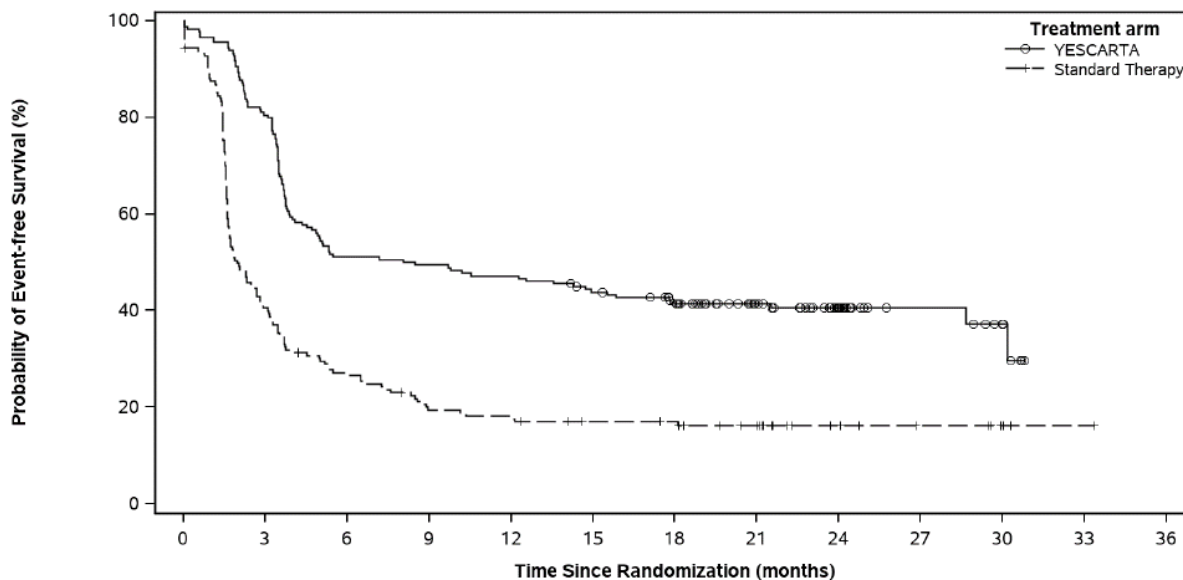
Table 7. Efficacy Results for ZUMA-7

Outcome^a	YESCARTA (N = 180)^g	Standard Therapy (N = 179)
Event-Free Survival^b		
Number of events, n (%)	108 (60)	144 (80)
Median, months [95% CI] ^c	8.3 [4.5, 15.8]	2.0 [1.6, 2.8]
Stratified hazard ratio [95% CI]	0.40 [0.31, 0.51]	
Stratified log-rank p-value	<0.0001	
Overall Survival^d		
Number of events, n (%)	82 (46)	95 (53)
Median OS, months [95% CI] ^c	NE (28.6, NE)	31.1 (17.1, NE)
Stratified hazard ratio [95% CI]	0.73 (0.54, 0.98)	
Stratified log-rank p-value ^e	0.0168	
Best Objective Response Rate, % [95% CI]		
	83 [77, 88]	50 [43, 58]
Difference in ORR, % [95% CI]	33 [23, 42]	
Stratified p-value ^f	<0.0001	
Complete remission rate, % [95% CI]	65 [58, 72]	32 [26, 40]
Partial remission rate, % [95% CI]	18 [13, 25]	18 [13, 24]
Progression-Free Survival		
Number of events, n (%)	93 (52)	81 (45)
Median, months [95% CI] ^c	14.9 [7.2, NE]	5.0 [3.4, 8.5]
Stratified hazard ratio [95% CI]	0.56 [0.41, 0.76]	

CI, confidence interval; NE, not estimable.

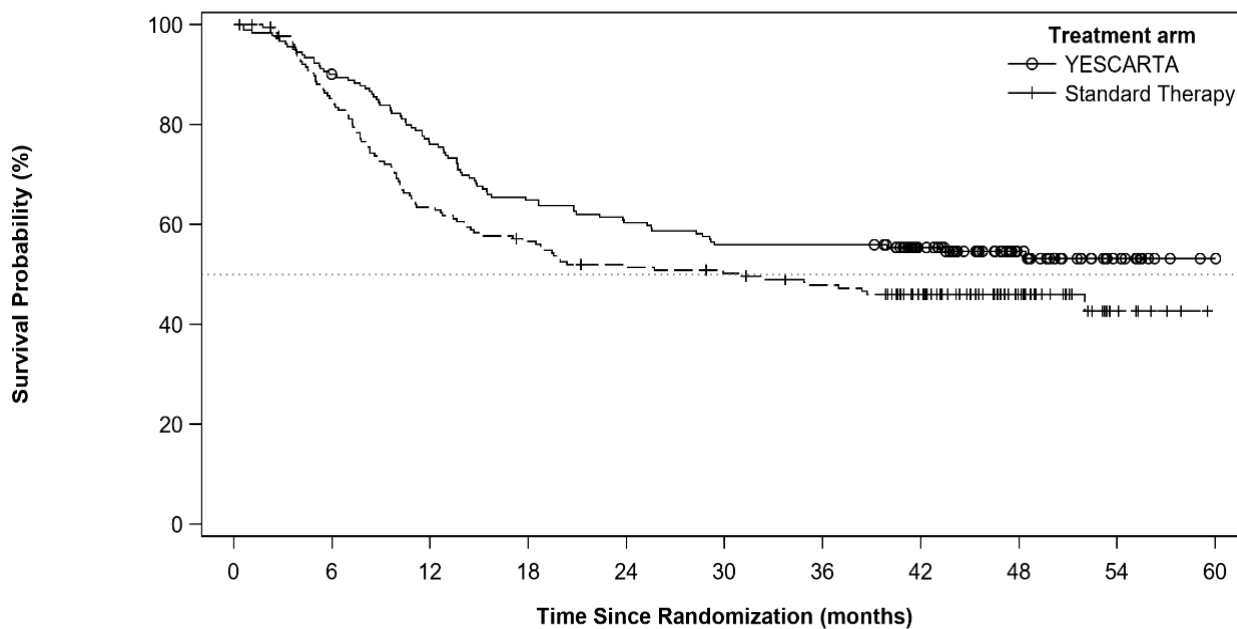
- Per the International Working Group Lugano Classification (Cheson 2014), as assessed by the independent review committee.
- EFS is defined as time from randomization to the earliest date of disease progression or relapse, best response of stable disease up to and including the Day 150 assessment, commencement of new lymphoma therapy, or death from any cause.
- Kaplan-Meier estimate.
- Overall survival was conducted at the time of the primary OS analysis
- p-value is compared with 0.0249, the one-sided efficacy boundary (significance level) for the primary OS analysis.
- Per Cochran-Mantel-Haenszel method. For all stratified analyses, stratification was based on response to first-line therapy (primary refractory, vs relapse within 6 months of first-line therapy vs relapse within > 6 but ≤ 12 months) and second-line age-adjusted International Prognostic Index.
- Two recipients of non-conformal product are included in the efficacy analysis.

Figure 1. Kaplan-Meier Curve of Event-Free Survival in ZUMA-7 (Primary EFS Analysis)



	Subjects at risk												
	0	3	6	9	12	15	18	21	24	27	30	33	36
YESCARTA	180	145	92	89	85	77	67	46	26	12	6	0	0
Standard Therapy	179	70	45	32	29	25	24	18	9	6	3	1	0

Figure 2. Kaplan-Meier Curve of Overall Survival in ZUMA-7 (Primary OS Analysis)



	Subjects at risk										
	0	6	12	18	24	30	36	42	48	54	60
YESCARTA	180	161	136	116	108	100	100	80	41	14	1
Standard Therapy	179	149	111	98	88	85	79	63	31	7	0

ZUMA-1

A single-arm, open-label, multicenter trial evaluated the efficacy of a single infusion of YESCARTA in adult patients with relapsed or refractory LBCL (ZUMA-1; NCT02348216). Eligible patients had refractory disease to the most recent therapy or relapse within 1 year after autologous hematopoietic stem cell transplantation (HSCT). The study excluded patients with prior allogeneic HSCT, any history of central nervous system lymphoma, ECOG performance status of 2 or greater, absolute lymphocyte count less than 100/ μ L, creatinine clearance less than 60 mL/min, hepatic transaminases more than 2.5 times the upper limit of normal, cardiac ejection fraction less than 50%, or active serious infection.

Following lymphodepleting chemotherapy, YESCARTA was administered as a single intravenous infusion at a target dose of 2×10^6 CAR-positive viable T cells/kg (maximum permitted dose: 2×10^8 cells). The lymphodepleting regimen consisted of cyclophosphamide 500 mg/m² intravenously and fludarabine 30 mg/m² intravenously, both given on the fifth, fourth, and third day before YESCARTA. Bridging chemotherapy between leukapheresis and lymphodepleting chemotherapy was not permitted. All patients were hospitalized for YESCARTA infusion and for a minimum of 7 days afterward.

Of 111 patients who underwent leukapheresis during Phase 2, 101 received YESCARTA. Of the patients treated, the median age was 58 years (range: 23 to 76 years), 67% were male, and 89% were white. Most (76%) had DLBCL, 16% had transformed follicular lymphoma, and 8% had primary mediastinal large B-cell lymphoma. The median number of prior therapies was 3 (range: 1 to 10), 77% of the patients had refractory disease to a second or greater line of therapy, and 21% had relapsed within 1 year of autologous HSCT.

One out of 111 patients did not receive the product due to manufacturing failure. Nine other patients were not treated, primarily due to progressive disease or serious adverse reactions following leukapheresis. The median time from leukapheresis to product delivery was 17 days (range: 14 to 51 days), and the median time from leukapheresis to infusion was 24 days (range: 16 to 73 days). The median dose was 2.0×10^6 CAR-positive viable T cells/kg (range: 1.1 to 2.2×10^6 cells/kg).

DLBCL in ZUMA-1 included patients with DLBCL not otherwise specified, other DLBCL subtypes, and HGBL based on the 2016 WHO-classification. Forty-seven patients were evaluable for MYC, BCL-2, and BCL-6 status. Thirty were found to have double expressor DLBCL (overexpression of both MYC and BCL-2 protein); 5 were found to have HGBL with MYC, BCL-2 or BCL-6 gene rearrangement (double- and triple-hit); and 2 were found to have HGBL not otherwise specified. Sixty-six patients were evaluable for cell-of-origin classifications (germinal center B-cell type [GCB] or activated B-cell type [ABC]). Of these, 49 patients had GCB-type and 17 patients had ABC-type.

The response rate in patients with relapsed or refractory LBCL is presented in Table 8. The median time to response was 1.0 months (range: 0.8 to 12.2 months). Response durations were longer in patients who achieved complete remission (CR), as compared to patients with a best response of partial remission (PR) (Table 9). Of the 55 patients who achieved CR, 17 initially had stable disease (7 patients) or PR (10 patients), with a median time to improvement of 2.1 months (range: 1.8 to 14.4 months).

Table 8. Response Rate in Patients with Relapsed or Refractory LBCL by Central Assessment

	Recipients of YESCARTA (N = 101)
Objective Response Rate (ORR)^a (95% CI)	75 (74%) (65%, 82%)
Complete Remission Rate (95% CI)	55 (54%) (44%, 64%)
Partial Remission Rate (95% CI)	20 (20%) (13%, 29%)

CI, confidence interval.

a. Per 2007 revised International Working Group criteria, as assessed by the independent review committee.

Table 9. Duration of Response (DOR) in Patients with Relapsed or Refractory LBCL by Central Assessment

	From N of 101
Number of Responders	75
DOR (Months)^a	
Median ^b	NE
(95% CI)	(10.9, NE)
Range ^c	0.0, 29.5+
DOR if Best Response is CR (Months)	
Median ^b	NE
(95% CI)	(NE, NE)
Range ^c	0.4, 29.5+
DOR if Best Response is PR (Months)	
Median ^b	2.1
(95% CI)	(1.3, 11.1)
Range ^c	0.0, 20.3+
Median Follow-up for DOR (Months)^{a, b}	22.9

CR, complete remission; DOR, duration of response; NE, not estimable; PR, partial remission.

- Among all responders. DOR is measured from the date of first objective response to the date of progression or death from relapse or toxicity.
- Kaplan-Meier estimate.
- A “+” sign indicates a censored value.

SCHOLAR-1

A retrospective, patient-level, pooled analysis of outcomes in refractory aggressive NHL (N = 636) was conducted (Crump et al., 2017) to provide confirmation of the prespecified control response rate of 20% and historical context for interpreting the ZUMA-1 results. The analysis included patients who had not responded (SD or PD) to their last line of therapy, or had relapsed within 12 months after ASCT. Response and survival after treatment with available standard-of-care therapy was evaluated. The ORR was 26% [95% CI (21, 31)] and the CR rate was 7% [95% CI (3, 15)], with a median OS of 6.3 months.

12 REFERENCES

- Lee DW et al (2014). Current concepts in the diagnosis and management of cytokine release syndrome. *Blood*. 2014 Jul 10; 124(2): 188-195.

13 HOW SUPPLIED/STORAGE AND HANDLING

YESCARTA is supplied in an infusion bag containing approximately 68 mL of frozen suspension of genetically modified autologous T cells in 5% DMSO and 2.5% albumin (human).

Each YESCARTA infusion bag is individually packed in a metal cassette. YESCARTA is stored in the vapor phase of liquid nitrogen and supplied in a liquid nitrogen dry shipper.

- Match the identity of the patient with the patient identifiers on the cassette and infusion bag upon receipt.
- Store YESCARTA frozen in the vapor phase of liquid nitrogen (less than or equal to minus 150°C).
- Thaw before using [see *Dosage and Administration (2)*].

14 SHELF LIFE

Refer to patient labels.

The stability of YESCARTA upon completion of thawing is up to 3 hours at room temperature (20°C to 25°C). However, YESCARTA infusion must begin within 30 minutes of thaw completion and the total YESCARTA infusion time must not exceed 30 minutes.

15 PATIENT COUNSELING INFORMATION

Advise the patient to read the HSA approved Patient Information Leaflet.

Ensure that patients understand the risk of manufacturing failure (< 1% in clinical trials). In case of a manufacturing failure, a second manufacturing of YESCARTA may be attempted. In addition, while the patient awaits the product, additional chemotherapy (not the lymphodepletion) may be necessary and may increase the risk of adverse events during the pre-infusion period.

Advise patients to seek immediate attention for any of the following:

- Cytokine Release Syndrome (CRS) - Signs or symptoms associated with CRS, including fever, chills, fatigue, tachycardia, nausea, hypoxia, and hypotension [*see Warnings and Precautions (5.1) and Adverse Reactions (6)*].
- Neurologic Toxicities - Signs or symptoms associated with neurologic events, including encephalopathy, seizures, changes in level of consciousness, speech disorders, tremors, and confusion [*see Warnings and Precautions (5.2) and Adverse Reactions (6)*].
- Serious Infections - Signs or symptoms associated with infection [*see Warnings and Precautions (5.5) and Adverse Reactions (6)*].
- Prolonged Cytopenia - Signs or symptoms associated with bone marrow suppression, including neutropenia, anemia, thrombocytopenia, or febrile neutropenia [*see Warnings and Precautions (5.6) and Adverse Reactions (6)*].

Advise patients of the need to:

- Refrain from driving or operating heavy or potentially dangerous machinery after YESCARTA infusion for at least 8 weeks after infusion [*see Warnings and Precautions (5.9)*].
- Have periodic monitoring of blood counts.

Product Owner:

Kite Pharma, Inc.
2400 Broadway
Santa Monica, CA 90404
USA

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